Volume 9, Number 3, 2007 © Mary Ann Liebert, Inc. DOI: 10.1089/ars.2006.1465

Forum Review

Involvement of Oxidative Stress in the Pathogenesis of Diabetes

HIDEAKI KANETO, NAOTO KATAKAMI, DAN KAWAMORI, TAKESHI MIYATSUKA, KEN'YA SAKAMOTO, TAKA-AKI MATSUOKA, MUNEHIDE MATSUHISA, and YOSHIMITSU YAMASAKI

ABSTRACT

Pancreatic β -cell failure is the common characteristic of type 1 and type 2 diabetes. Type 1 diabetes is induced by pancreatic β -cell destruction, which is mediated by an autoimmune mechanism and consequent inflammatory process. Various inflammatory cytokines and oxidative stress produced by islet-infiltrating immune cells have been proposed to play an important role in mediating the destruction of β cells. The JNK pathway is also activated by such cytokines and oxidative stress and is involved in β -cell destruction. Type 2 diabetes is the most prevalent and serious metabolic disease affecting people all over the world. Pancreatic β -cell dysfunction and insulin resistance are the hallmark of type 2 diabetes. Once hyperglycemia becomes apparent, β -cell function gradually deteriorates, and insulin resistance is aggravated. This process is called "glucose toxicity." Under such conditions, oxidative stress is provoked, and the JNK pathway is activated, which is likely involved in pancreatic β -cell dysfunction and insulin resistance. In addition, oxidative stress and activation of the JNK pathway are involved in the progression of atherosclerosis, which is often observed under diabetic conditions. Taken together, it is likely that oxidative stress and subsequent activation of the JNK pathway are involved in the pathogenesis of type 1 and type 2 diabetes. Antioxid. Redox Signal. 9, 355–366.

INTRODUCTION

TAILURE OF PANCREATIC β cells is the common characteristic of type 1 and type 2 diabetes. Type 1 diabetes mellitus is induced by destruction of pancreatic β cells, which is mediated by an autoimmune mechanism and the consequent inflammatory process. Various inflammatory cytokines and oxidative stress are produced during this process, which has been proposed to play an important role in mediating β -cell destruction. Type 2 diabetes is the most prevalent and serious metabolic disease, and β -cell dysfunction and insulin resistance are the hallmark of type 2 diabetes. Under diabetic conditions, chronic hyperglycemia gradually deteriorates β -cell function and aggravates insulin resistance. This process is called "glucose toxicity." Oxidative stress is provoked during this process and is likely involved in β -cell dysfunction and insulin resistance. Here we show that oxidative stress and

subsequent activation of the JNK pathway are involved in the pathogenesis of both type 1 and type 2 diabetes.

OXIDATIVE STRESS AND β-CELL DESTRUCTION IN TYPE 1 DIABETES

Type 1 diabetes is mediated by an autoimmune mechanism or inflammatory process that is characterized by destruction of pancreatic β cells (26, 84, 105). The initial event in an immune response is the uptake of antigen by antigen-presenting cells (APCs) such as macrophages, and the cooperation of CD4+ and CD8+ T cells is crucial for islet infiltration and destruction of β cells (25, 134). CD4+ Th1/Th2 balance is also critical for the development and resolution of immune responses. CD4+ Th1 T-cells produce interleukin 2 (IL-2) and interferon γ (IFN- γ), which leads to activation of CD8+ T

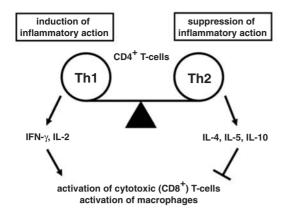


FIG. 1. Effects of Th1/Th2 balance of CD4+ T cells on the activation of CD8+ T cells and macrophages.

cells and macrophages. In contrast, CD4⁺ Th2 T cells secrete IL-4, IL-5, and IL-10, which leads to suppression of inflammatory action (Fig. 1). Activation of CD8⁺ T cells responds to class I molecules in β cells themselves, which is a critical step in the process of β -cell destruction (Fig. 2). Activated macrophages secrete various inflammatory cytokines and reactive oxygen species (ROS), which are also involved in the process of β -cell destruction (see Fig. 2).

Various inflammatory cytokines and ROS have been proposed to play an important role in β -cell destruction. Activated macrophages produce inflammatory cytokines, such as IL-1 β and tumor necrosis factor α (TNF- α), and ROS, such as superoxide anion, hydrogen peroxide, and nitric oxide (NO) (see Fig. 2). Among various inflammatory cytokines, IL-1 β has been proposed to play an important role in mediating β -cell destruction (4, 6, 18, 19, 20, 28, 57, 120, 122). The deleterious effects of IL-1 β have been proposed to involve generation of NO and inhibition of mitochondrial function. Indeed, IL-1 β -induced production of nitrite and iron-nitrosyl complex in β cells was confirmed using electron paramagnetic resonance (18).

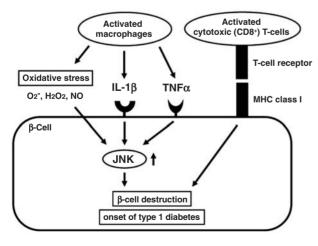


FIG. 2. Involvement of oxidative stress and the JNK pathway in pancreatic β -cell destruction in type 1 diabetes.

NO seems to be an important inducer of necrosis and perhaps even apoptosis, especially in rodent islets. Because pancreatic B cells contain very low levels of antioxidant enzymes, they may be more susceptible to the toxic actions of cytokines (27, 28). Overexpression of antioxidant enzymes protected in several insulin-producing tumor cell lines against the toxic effects of both oxidative stress and inflammatory cytokines (10, 17, 27, 44, 47, 74, 75, 78, 80, 127). For example, overexpression of the mitochondrial form of superoxide dismutase (Mn-SOD) protected β cells against oxidative stress and cytokines (10, 17, 44, 80). Targeted overexpression of the cytosolic SOD (Cu/Zn-SOD) in B cells protected mice from autoimmune and low-dose streptozotocin-induced diabetes (74, 75). Furthermore, it was shown that a SOD mimetic with longer half-life was effective to protect mice against the onset of type 1 diabetes (102). These data suggest that oxidative stress is involved in β-cell destruction and the onset of type 1 diabetes.

THE JNK PATHWAY AND β-CELL DESTRUCTION IN TYPE 1 DIABETES

Studies on insulin-secreting cells and primary β cells have revealed that IL-1 β is a potent activator of the JNK pathway (76, 83, 133). More-convincing results have been obtained on the role of JNK in controlling IL-1 β -mediated apoptosis. Transfection experiments and the use of cell-permeable peptide inhibitors demonstrated that inhibiting the JNK pathway confers protection against apoptosis induced by IL-1 β in insulin-secreting cells (3, 12, 13). Given the fact that TNF- α and IFN- γ strongly potentiate the cytotoxic effects of IL-1 β on β cells, it is of note that these two cytokines synergistically augment IL-1 β -induced signaling via the JNK pathway in rat islets (5). This observation may provide at least a partial explanation at the signaling level for the synergistic toxic effects of cytokines on β cells.

It has been reported recently that JNK2 plays an important role in type 1 diabetes that is caused by autoimmune destruction of β cells (53). Studies of nonobese diabetic mice demonstrated that disruption of the JNK2 protein kinase decreased destructive insulitis and reduced disease progression to diabetes. CD4+ T cells from JNK2-deficient nonobese diabetic mice produced less IFN- γ but significantly increased amounts of IL-4 and IL-5, indicating polarization toward the Th2 phenotype. This role of JNK2 to control the Th1/Th2 balance of the immune response represents a mechanism of protection against autoimmune diabetes. These results indicate that JNK2 protein kinase plays an important role in the onset of type 1 diabetes.

OXIDATIVE STRESS AND β -CELL DYSFUNCTION IN TYPE 2 DIABETES

It has been suggested that although exposure of β cells to a high glucose concentration for relatively short periods stimulates insulin gene expression, prolonged exposure has adverse effects on various β -cell functions. Chronic hyper-

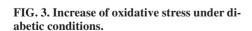
glycemia is a cause of impairment of insulin biosynthesis and secretion; once hyperglycemia becomes apparent, β-cell function gradually deteriorates. This process is called "β-cell glucose toxicity," which is often observed under diabetic conditions (54, 93, 103, 104, 115, 131). In the diabetic state, hyperglycemia per se and subsequent production of oxidative stress decrease insulin gene expression and secretion (9, 21, 31, 35, 39, 49, 54, 56, 58, 59, 63, 82, 85, 108, 109, 123, 124). It was previously shown that the loss of insulin gene expression is accompanied by decreased expression and/or DNAbinding activities of transcription factors pancreatic and duodenal homeobox-1 (PDX-1) (49, 58, 82, 85, 93, 104, 123) and RIPE3b1-binding protein (which was recently identified as MafA) (39, 104, 115, 131), After long-term exposure to a high glucose concentration, expression and/or DNA-binding activities of these two transcription factors are reduced. It is noted here that PDX-1, also known as IDX-1/STF-1/IPF1, plays a crucial role in pancreas development, \u03b3-cell differentiation, and induction of surrogate \(\beta \) cells (2, 32, 45, 55, 79, 89, 90, 91, 94, 97, 99). MafA is a recently isolated βcell-specific transcription factor that functions as a potent activator of insulin gene transcription (62, 66, 86, 87, 100, 139).

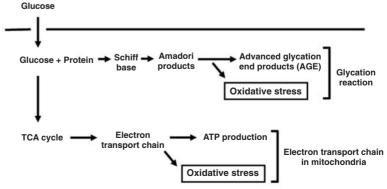
Under diabetic conditions, oxidative stress is provoked (9, 21, 49) and involved in the β-cell glucose toxicity found in diabetes (31, 35, 39, 54, 56, 58, 59, 63, 82, 85, 108, 109, 123, 124). B Cells express GLUT2, a high-Km glucose transporter, and thereby display highly efficient glucose uptake when exposed to a high glucose concentration. Indeed, it was shown that expression of oxidative stress markers 8-hydroxy-2'-deoxyguanosine (8-OHdG) and 4-hydroxy-2,3-nonenal (4-HNE) were increased in islets under diabetic conditions (35, 49). In addition, β cells are rather vulnerable to oxidative stress because of the relatively low expression of antioxidant enzymes such as catalase and glutathione peroxidase (15). Thus, it is likely that oxidative stress is involved in β -cell deterioration found in diabetes. Several sources of ROS exist in cells, such as the nonenzymatic glycosylation reaction (85), and the electron-transport chain in mitochondria (112) (Fig. 3).

It was shown that when β -cell–derived cell lines or rat isolated islets were exposed to oxidative stress, insulin gene promoter activity and mRNA expression were suppressed (56, 58, 59, 85, 123). In addition, when β -cell–derived cell lines

or rat isolated islets were exposed to oxidative stress, binding of PDX-1 to the insulin gene promoter was markedly reduced. Furthermore, it was shown that the decrease of insulin gene expression after prolonged exposure to a high glucose concentration was prevented by treatment with antioxidants (39, 58, 123, 124). Reduction of expression and/or DNAbinding activities of PDX-1 and MafA by long-term exposure to high glucose was also prevented by an antioxidant treatment. These results suggest that chronic hyperglycemia suppresses insulin biosynthesis and secretion by provoking oxidative stress, accompanied by reduction of expression and/or DNA-binding activities of two important pancreatic transcription factors, PDX-1 and MafA. Therefore, it is likely that the alteration of such transcription factors explains, at least in part, the suppression of insulin biosynthesis and secretion, and thus are involved in β -cell glucose toxicity.

Next, to evaluate the potential usefulness of antioxidants in treatment for type 2 diabetes, obese diabetic C57BL/KsJdb/db mice were treated with antioxidants (N-acetyl-L-cysteine plus vitamin C and E) (58). The antioxidant treatment retained glucose-stimulated insulin secretion and moderately ameliorated glucose tolerance. β-Cell mass was significantly larger in the mice treated with the antioxidants. Insulin content and insulin mRNA levels were also preserved by the antioxidant treatment. Furthermore, PDX-1 expression was more clearly visible in the nuclei of islet cells after the antioxidant treatment (58). Similar effects were observed with Zucker diabetic fatty rats, another model animal for type 2 diabetes (123). Taken together, these data indicate that antioxidant treatment can protect β cells against glucose toxicity. In addition, as a step to clinical trial of antioxidant for type 2 diabetes, we examined the possible antidiabetic effects of probucol, an antioxidant widely used as an anti-hyperlipidemic agent, on preservation of \(\beta\)-cell function in diabetic C57BL/KsJ-db/db mice (35). Immunostaining for oxidative stress markers such as 4-hydroxy-2-nonenal (HNE)-modified proteins and heme oxygenase-1 revealed that probucol treatment decreased ROS in B cells of diabetic mice. Probucol treatment also preserved β-cell mass, insulin content, and glucose-stimulated insulin secretion, leading to improvement of glucose tolerance (35). These data suggest potential usefulness of antioxidants for diabetes and provide further support for the implication of oxidative stress in β-cell glucose toxicity found in diabetes.





It is noted here that lipotoxicity as well as glucose toxicity is involved in the deterioration of β -cell function found in type 2 diabetes (104, 116, 117). Indeed, it has been shown that deranged lipid metabolism in β cells, most typically represented by accumulation of intracellular triglycerides, causes β -cell damage through induction of iNOS and excess nitric oxide (NO) generation in Zucker diabetic fatty rats (116). Because serum free fatty acid levels and islet triglyceride content were decreased by probucol treatment, decrease of lipotoxicity might have also contributed to the preservation of β -cell function and amelioration of glucose tolerance after probucol treatment (35).

THE JNK PATHWAY AND β-CELL DYSFUNCTION IN TYPE 2 DIABETES

It has been suggested that activation of the c-Jun N-terminal kinase (JNK) pathway is involved in pancreatic β-cell dysfunction found in diabetes. It was reported that activation of the JNK pathway is involved in reduction of insulin gene expression by oxidative stress and that suppression of the JNK pathway can protect β cells from oxidative stress (60). When isolated rat islets were exposed to oxidative stress, the JNK pathway was activated, preceding the decrease of insulin gene expression. Adenoviral overexpression of dominantnegative type JNK1 (DN-JNK) protected insulin gene expression and secretion from oxidative stress. Moreover, wild-type JNK1 (WT-JNK) overexpression suppressed both insulin gene expression and secretion (60). These results were correlated with change in the binding of the important transcription factor PDX-1 to the insulin promoter. Adenoviral overexpression of DN-JNK preserved PDX-1 DNA-binding activity in the face of oxidative stress, whereas WT-JNK overexpression decreased PDX-1 DNA binding activity (60). Thus, it is likely that JNK-mediated suppression of PDX-1 DNA binding activity accounts for some of the suppression of insulin gene transcription and of β-cell function, which fits with the phenomenon that PDX-1 DNA-binding activity is decreased in association with reduction of insulin gene transcription after prolonged exposure to a high glucose concentration. Taken together, it is likely that activation of the JNK pathway leads to decreased PDX-1 activity and consequent suppression of insulin gene transcription found in the diabetic state (Fig. 4).

As a potential mechanism for JNK-mediated PDX-1 inactivation, it was recently reported that PDX-1 is translocated from the nuclei to the cytoplasm in response to oxidative stress. When oxidative stress was charged on β-cell-derived HIT cells, both intrinsically expressed PDX-1 and exogenously introduced green fluorescent protein (GFP)-tagged PDX-1 moved from the nuclei to the cytoplasm (77). Addition of DN-JNK inhibited the oxidative stress-induced PDX-1 translocation, suggesting an essential role of JNK in mediating the phenomenon. Whereas the nuclear localization signal (NLS) in PDX-1 was not affected by oxidative stress, leptomycin B, a specific inhibitor of the classic, leucine-rich nuclear export signal (NES), inhibited nucleocytoplasmic translocation of PDX-1 induced by oxidative stress. Indeed,

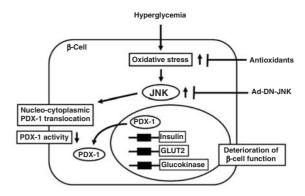


FIG. 4. Involvement of oxidative stress and the JNK pathway in pancreatic β -cell glucose toxicity found in type 2 diabetes.

we identified an NES at position 82–94 of the mouse PDX-1 protein (77). Taken together, it is likely that oxidative stress induces nucleocytoplasmic translocation of PDX-1 through activation of the JNK pathway, which leads to reduction of its DNA-binding activity and suppression of insulin biosynthesis (see Fig. 4).

Furthermore, we recently showed that the forkhead transcription factor Foxo1 plays a role as a mediator between the JNK pathway and PDX-1 (78). In β-cell-derived cell line HIT-T15, Foxo1 changed its intracellular localization from the cytoplasm to the nucleus under oxidative stress conditions. In contrast to Foxo1, the nuclear expression of PDX-1 was decreased, and its cytoplasmic distribution was increased by oxidative stress. The overexpression of JNK also induced the nuclear localization of Foxo1, but in contrast, suppression of the JNK pathway reduced the oxidative stress-induced nuclear localization of Foxo1, suggesting an involvement of the JNK pathway in Foxo1 translocation (78). In addition, oxidative stress or activation of the JNK pathway decreased Akt phosphorylation in HIT cells, leading to the decreased phosphorylation of Foxo1 after nuclear localization. Furthermore, adenoviral Foxo1 overexpression reduced the nuclear expression of PDX-1, whereas repression of Foxo1 by Foxo1specific small interfering RNA retained the nuclear expression of PDX-1 under oxidative stress conditions (78). Taken together, oxidative stress and subsequent activation of the JNK pathway induce nuclear translocation of Foxo1 through the modification of the insulin signaling in b cells, which leads to the nucleocytoplasmic translocation of PDX-1 and reduction of its DNA-binding activity (Fig. 5). Furthermore, it has been shown very recently that the protein kinase MST1 is activated by oxidative stress, which leads to facilitation of Foxo1 translocation from cytoplasm to nuclei (77). Therefore, it is also possible that oxidative stress triggers Foxo1 translocation from cytoplasm to nuclei, independent of Akt activity or Akt-mediated phosphorylation status of Foxo1.

In addition, the significance of JNK in the development of diabetes comes from the result of a genetic analysis in humans. Although islet-brain-1 (IB1) was known to regulate the JNK pathway (11, 24), it was reported that a missense mutation within the IB1-encoding MAPKIP1 gene (S59N) is associated with a late-onset type 2 diabetes (130). Thus, it is

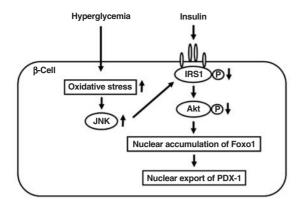


FIG. 5. Involvement of Foxo1 and insulin signaling in pancreatic β -cell dysfunction found in type 2 diabetes.

likely that activation of the JNK pathway is involved in deterioration of β -cell function found in type 2 diabetes.

OXIDATIVE STRESS AND INSULIN RESISTANCE IN TYPE 2 DIABETES

The hallmark of type 2 diabetes is insulin resistance as well as pancreatic β-cell dysfunction. Under diabetic conditions, various insulin target tissues such as liver, muscle, and fat become resistant to insulin. The pathophysiology of insulin resistance involves a complex network of insulin signaling pathways. After insulin binds to insulin receptor on the cell surface, insulin receptor and its substrates are phosphorylated, which leads to activation of various insulin signaling pathways. It has been shown that oxidative stress is involved in progression of insulin resistance as well as pancreatic β-cell dysfunction (30). It was previously reported that oxidative stress disrupted insulin-induced cellular redistribution of insulin receptor substrate-1 (IRS-1) and phosphatidylinositol 3-kinase (PI 3-K) and thus impaired insulininduced GLUT4 translocation in the 3T3-L1 adipocyte (111, 128). It was also reported that treatment with antioxidants (N-acetyl-L-cysteine and taurine) prevented hyperglycemiainduced insulin resistance in vivo (38). Furthermore, in patients with type 2 diabetes, both short- and long-term administration of α -lipoic acid, an antioxidant, improved insulin resistance, as measured by both the euglycemic-hyperinsulinemic clamp and the Bergman minimal model (52, 72). These data indicate that oxidative stress is involved in the progression of insulin resistance.

THE JNK PATHWAY AND INSULIN RESISTANCE IN TYPE 2 DIABETES

In diabetic patients, hyperglycemia increased oxidative stress, which leads to activation of the JNK pathway. In addition, under diabetic conditions, various inflammatory cytokines and intracellular lipid accumulation also lead to activation of the JNK pathway. It has been suggested that

activation of the JNK pathway is involved in insulin resistance as well as the pancreatic β-cell dysfunction found in diabetes (46, 132). It was reported that the JNK pathway is abnormally activated in the liver, muscle, and adipose tissue in obese type 2 diabetic mice and that insulin resistance in obese type 2 diabetic mice is substantially reduced in mice homozygous for a targeted mutation in the JNK1 gene (JNK-KO mice) (43). When the JNK-KO mice were placed on a highfat/high-caloric diet, obese wild-type mice developed mild hyperglycemia compared with lean wild-type mice. In contrast, blood glucose levels in obese JNK-KO mice was significantly lower compared with those in obese wild-type mice. Intraperitoneal insulin tolerance tests showed that the hypoglycemic response to insulin in obese wild-type mice was lower compared with that in obese JNK-KO mice. Intraperitoneal glucose tolerance tests revealed a higher degree of hyperglycemia in obese wild-type mice than in obese JNK-KO mice. These results indicate that the JNK-KO mice are protected from the development of dietary obesity-induced insulin resistance.

Furthermore, targeted mutations in JNK were introduced in genetically obese mice (ob/ob) (43). Blood glucose levels in ob/ob-JNK-KO mice were lower compared with those in ob/ob wild-type mice, and the ob/ob wild-type mice displayed a severe and progressive hyperinsulinemia. Thus, JNK deficiency can provide partial resistance against obesity, hyperglycemia, and hyperinsulinemia in both genetic and dietary models of diabetes. Such improvement of insulin resistance and glucose tolerance might be accounted for by reduced body-weight gain by JNK deficiency in mice. Taken together, obese type 2 diabetes is associated with activation of the JNK pathway, and the absence of JNK results in substantial protection from obesity-induced insulin resistance. These results strongly suggest that JNK plays a crucial role in progression of insulin resistance found in type 2 diabetes (Fig. 6). It is noted here that, of the three isozymes of JNK (JNK1, JNK2, and JNK3), only JNK1 has been shown to be implicated in the development of insulin resistance (43).

It was also reported that overexpression of dominantnegative (DN) type JNK1 (Ad-DN-JNK) in the liver of obese diabetic C57BL/KsJ-db/db mice dramatically improved insulin resistance and markedly decreased blood glucose levels (43). In an intraperitoneal insulin tolerance test, the hypoglycemic response to insulin was larger in Ad-DN-JNK-treated db/db mice. Furthermore, in the euglycemic

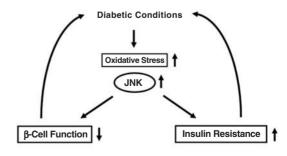


FIG. 6. Involvement of oxidative stress and the JNK pathway in the pathogenesis of type 2 diabetes.

hyperinsulinemic clamp test, the glucose infusion rate (GIR) in Ad-DN-JNK- treated mice was higher than that in Ad-GFP-treated mice, indicating that suppression of the JNK pathway in the liver reduces insulin resistance and thus ameliorates glucose tolerance in the db/db mice. Furthermore, hepatic glucose production (HGP) was significantly lower in Ad-DN-JNK-treated mice. In contrast, no difference was found in the glucose disappearance rate (Rd) between these two groups (96). These results indicate that reduction of insulin resistance and amelioration of glucose tolerance by DN-JNK overexpression are mainly due to suppression of hepatic glucose production. It has been reported that serine phosphorylation of insulin receptor substrate-1 (IRS-1) inhibits insulin-stimulated tyrosine phosphorylation of IRS-1, leading to an increase in insulin resistance (1). IRS-1 serine 307 phosphorylation was markedly decreased in Ad-DN-JNK-treated mice. An increase in IRS-1 tyrosine and Akt serine 473 phosphorylation was also observed in Ad-DN-JNK-treated mice (96). Therefore, an increase in IRS-1 serine phosphorylation may be closely associated with the development of insulin resistance induced by JNK overexpression. These results indicate that suppression of the JNK pathway enhances insulin signaling, which leads to amelioration of glucose tolerance. Taken together, these findings suggest that suppression of the JNK pathway in the liver exerts greatly beneficial effects on insulin-resistance status and glucose tolerance in both genetic and dietary models of diabetes.

Protein transduction domains (PTDs), such as the small PTD from the TAT protein of human immunodeficiency virus (HIV-1), the VP22 protein of Herpes simplex virus, and the third α-helix of the homeodomain of Antennapedia, a Drosophila transcription factor, are known to allow various proteins and peptides to be efficiently delivered into cells through the plasma membrane, and thus increasing interest has been expressed in their potential usefulness for the delivery of bioactive proteins and peptides into cells (1, 29, 34, 95, 98, 110, 114). It was recently reported that the cell-permeable JNK inhibitory peptide is effective for the treatment of diabetes. This peptide is derived from the JNK-binding domain of JNK-interacting protein-1 (JIP-1) and has been reported to function as a dominant inhibitor of the JNK pathway (13). To convert the minimal JNK-binding domain into a bioactive cell-permeable compound, a 20-amino-acid sequence derived from the JNK-binding domain of JIP-1 was covalently linked to a 10-amino-acid carrier peptide derived from the HIV-TAT sequence; then to monitor peptide delivery, this JNK inhibitory peptide was further conjugated with fluorescein isothiocyanate (FITC). When this peptide was injected intraperitoneally to C57BL/KsJ-db/db obese diabetic mice, the FITC-conjugated peptide showed fluorescence signals in insulin target organs (liver, fat, muscle) and in insulin-secreting tissue (pancreatic islets) (61). In insulin tolerance tests, reduction of blood glucose levels in response to injected insulin was much larger in JNK inhibitory peptide-treated mice (61). Furthermore, in the euglycemic hyperinsulinemic clamp test, the steady-state GIR in JNK inhibitory peptide-treated mice was significantly higher than that in untreated mice, indicating that JNK inhibitory peptide reduces insulin resistance in the db/db mice.

Endogenous hepatic glucose production (HGP) and glucose disappearance rate (Rd) in the JNK inhibitory peptidetreated mice also was evaluated. It is noted that Rd reflects glucose utilization in the peripheral tissues. HGP in JNK inhibitory peptide-treated mice was significantly lower than that in untreated mice. In addition, Rd in JNK inhibitory peptide-treated mice was significantly higher than that in untreated mice (61). These results indicate that JNK inhibitory peptide treatment reduces insulin resistance through decreasing HGP and increasing Rd. IRS-1 serine 307 phosphorylation was decreased in JNK inhibitory peptide-treated mice. An increase of IRS-1 tyrosine phosphorylation was observed in the peptide-treated mice. Concomitantly, glucose tolerance was also ameliorated in JNK inhibitory peptide-treated mice. In conclusion, suppression of the JNK pathway improves insulin resistance and ameliorates glucose intolerance, indicating that the JNK pathway plays a crucial role and could be a potential therapeutic target for diabetes (see Fig. 6).

OXIDATIVE STRESS AND ATHEROSCLEROSIS

Atherosclerosis is often observed in subjects with various metabolic diseases such as diabetes, hypertension, and hyperlipidemia. Among various risk factors, diabetes is thought to be one of the most important risk factors that facilitate the progression of atherosclerosis. Atherosclerosis is often observed as a macroangiopathy under diabetic conditions. We reported that increase of intima-media thickness (IMT) in the carotid artery, an index of the progression of atherosclerosis, is often observed in patients with diabetes (69, 135, 136) and that the progression of IMT is attenuated after treatment with various medicines for diabetes (65, 137). It is well known that hyperglycemia per se found under diabetic conditions facilitates the progression of atherosclerosis. Furthermore, hyperinsulinemia, which is often observed in subjects with insulin resistance, is also likely involved in the progression of atherosclerosis (Fig. 7).

Endothelial dysfunction is likely to be one of the earliest key events in atherosclerosis (14, 88). It has been thought that oxidative stress is involved in the deterioration of endothelial function, which is accompanied by inactivation of endothelial nitric oxide synthase (eNOS) and decrease of available nitric oxide (NO). Oxidative stress also induces expression of adhesion molecules such as intercellular adhesion molecule-1 (ICAM-1) and vascular adhesion molecule-1 (VCAM-1), and receptors of oxidized lipid particles such as leptin-like oxidized LDL receptor (LOX-1). The expression of endothelial cell-surface receptors facilitates inflammatory cell recruitment and lipid deposition in the intimal layer. The subsequent ingestion of excess oxidized LDL particles by macrophages and monocytes results in inflammatory cytokine and growth factor release. Excessive ROS formation in diabetes further facilitates endothelial dysfunction, accompanied by inactivation of eNOS and decrease of NO (15). Indeed, it was reported that exposure of endothelial cells to a high glucose concentration inhibited eNOS expression and diminished NO bioavailability (16).

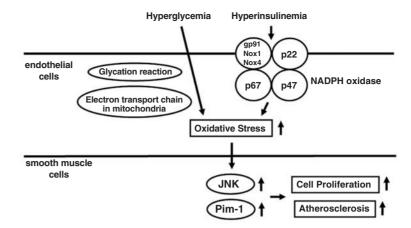


FIG. 7. Involvement of oxidative stress and the JNK pathway in the progression of atherosclerosis.

Proliferation of vascular smooth muscle cells (VSMCs) is also a key step in the development of atherosclerosis. It has been suggested that reactive oxygen species (ROS) and various growth factors are involved in the VSMC proliferation and the development of atherosclerosis (81). Induction of ROS is often observed in vivo under certain conditions such as diabetes, hypertension, hypercholesterolemia, obesity, and smoking, all of which are risk factors of atherosclerosis. Clinical mass studies have also provided support for the significance of ROS in the development of atherosclerosis (118, 126). Induction of ROS causes an increase in peroxidized lipids and thus enhances the foam-cell formation of macrophages and promotes migration of VSMCs. In addition, ROS directly stimulate DNA synthesis and accelerate VSMC proliferation, which plays a key role in the development of atherosclerosis. It has also been demonstrated that ROS regulate expression of various growth factors such as heparin-binding epidermal growth factor-like growth factor and insulin-like growth factor I and several growth-related proto-oncogenes such as c-Myc, c-Fos, and c-Jun (23, 106). Furthermore, using the suppression subtractive hybridization technique, we previously identified that ROS regulate expression of various other factors such as fibronectin, p105 co-activator, and ECA39 in VSMCs, all of which are likely involved in the progression of atherosclerosis (113). Taken together, it is likely that oxidative stress is involved in the VSMC proliferation and development of atherosclerosis through various pathways.

It has been shown that membrane-bound NADPH oxidase is the major source of ROS in the vasculature (40, 92) and that NADPH oxidase-derived ROS play a critical role in the development of atherosclerosis. NADPH oxidase is composed of the membrane-bound subunits gp91 phox (Nox2)/ Nox1/Nox4, and p22 phox, and the catalytic site of the oxidase and cytosolic components p47 phox and p67 phox. In vascular cells, such as endothelial and smooth muscle cells, Nox 1 and Nox 4, rather than gp91 phox, are abundantly expressed. NADPH oxidase is activated by various factors such as angiotensin II, thrombin, platelet-derived growth factor, and tumor necrosis factor- α (22, 101, 121, 129). It has been reported that mice lacking p47 phox, which is an important component for NADPH oxidase, have lower levels of aortic ROS production compared with wild-type mice, and, when in a hypercholesterolemic apolipoprotein E-deficient [apoE

(-/-)] background, had significantly fewer lesions in their descending aortas compared with apoE (-/-) mice (8). NADPH oxidase-derived ROS also play a crucial role in atherosclerosis in human. It has been reported that ROS production in atherosclerotic human coronary arteries is associated with NADPH oxidase subunit p22 phox (7) and recently that phagocytic NADPH oxidase overactivity is involved in oxidative stress and atherosclerosis in metabolic syndrome patients and that hyperinsulinemia may contribute to oxidative stress in metabolic syndrome patients through activation of NADPH oxidase (33). In addition, it was shown that high glucose stimulates ROS production through the activation of NADPH oxidase (42, 50) and that the p22 phox was significantly increased in rat and human diabetic arteries (37, 71). This activation of p22 phox may contribute to the acceleration of atherosclerosis in patients with diabetes. The role of p22 phox in the progression of atherosclerosis was also shown in type 2 diabetes patients. We previously reported that in type 2 diabetes subjects, the C242T polymorphism of the p22 phox gene, an essential component of NADPH oxidase in the vasculature, was closely associated with IMT of the carotid artery, an index of the progression of atherosclerosis (41). Average IMT in the diabetes patients with the CC genotype was significantly higher compared with those with the TC+TT genotypes, despite no difference in the risk factors. It is noted here that the presence of 242T allele is know to be associated with significantly reduced vascular NADPH oxidase activity (36). Furthermore, in stepwise multiple regression analysis, the p22 phox CC genotype was an independent risk factor for increased IMT in the diabetic subjects (41). These results show that the C242T mutation in the p22 phox gene is associated with progression of atherosclerosis in the diabetes patients and further strengthen the important role of NADPH oxidase in the progression of atherosclerosis.

Although various molecules and kinases are thought to be activated by oxidative stress in VSMCs, we recently reported that Pim-1, a proto-oncogene that encodes a serine/threonine kinase, is induced by oxidative stress and thus is likely involved in the progression of atherosclerosis (64). Pim-1 was substantially induced in neointimal VSMCs of balloon-injured rat carotid arteries, and *in vivo* infection with a dominant-negative Pim-1–expressing adenovirus (Ad-DN-Pim-1) markedly suppressed neointima formation

and cell-cycle progression in the balloon-injured arteries (64). In cultured VSMCs, ROS-stimulated cell-cycle progression and DNA synthesis were suppressed by DN-Pim-1 overexpression. Furthermore, Pim-1-producing cells were observed predominantly in the thickened intima of human thoracic aortas and coronary arteries (64). These findings suggest that oxidative stress and consequent induction of Pim-1 expression play a critical role in the progression of atherosclerosis (see Fig. 7).

THE JNK PATHWAY AND ATHEROSCLEROSIS

Although the JNK pathway is known to be activated by oxidative stress in VSMCs (138), oxidative stress and subsequent activation of the JNK pathway are likely involved in the progression of atherosclerosis. It is known that the JNK pathway is commonly activated by vascular remodeling-related molecules and plays a central role in the initiation of cellular responses, including cellular gene expression, growth, migration, or apoptosis. It was previously reported that the JNK pathway is activated in balloon-injured arteries (48, 70, 73). In vivo transfection of DN-JNK significantly suppressed activation of the JNK pathway and reduced VSMC proliferation in a balloon-injury model (51). Neointimal formation after balloon injury was also prevented by DN-JNK overexpression. Bromodeoxyuridine labeling index and total cell-counting analysis showed that DN-JNK remarkably suppressed VSMC proliferation in both the intima and the media after injury. In contrast, gene transfer of wild-type JNK (WT-JNK) significantly enhanced neointimal hyperplasia after balloon injury. Taken together, activation of the JNK pathway triggers VSMC proliferation, leading to neointimal formation, and the JNK pathway could be a new therapeutic target for atherosclerosis (see Fig. 7).

Furthermore, the role of JNK in atherosclerotic plaque formation in vivo was examined using atherosclerosis-prone apolipoprotein E knockout mice (ApoE (-/-) mice). Activation of the JNK pathway was closely correlated with the presence of clearly established plaques in ApoE (-/-) mice with a high-cholesterol diet. It was recently reported that atherosclerosis-prone ApoE (-/-) mice simultaneously lacking JNK2 [ApoE (-/-), JNK2 (-/-) mice], but not ApoE (-/-), JNK1 (-/-) mice, developed less atherosclerosis compared with ApoE (-/-) mice (107). Pharmacologic inhibition of the JNK activity also efficiently reduced plaque formation. Macrophages lacking JNK2 displayed suppressed foam cell formation caused by defective uptake and degradation of modified lipoproteins and showed increased amounts of the modified lipoprotein-binding and -internalizing scavenger receptor A (SR-A). Macrophage-restricted deletion of JNK2 was sufficient to decrease atherogenesis (107). These data suggest that JNK2-dependent phosphorylation of SR-A promotes uptake of lipids in macrophages, and thereby regulates foam cell formation. These data also further strengthen the significance of the JNK pathway in the progression of atherosclerosis (see Fig. 7).

CONCLUDING REMARKS

Oxidative stress and subsequent activation of the JNK pathway are involved in the pathogenesis of type 1 and type 2 diabetes. In the onset of type 1 diabetes, pancreatic β cells are relatively immediately destroyed by large amounts of ROS, various inflammatory cytokines, and activation of the stress signaling. Suppression of oxidative stress or the JNK pathway in β cells of type 1 diabetic mice suppresses the onset of type 1 diabetes. Oxidative stress is induced and the JNK pathway is activated under hyperglycemic conditions, which is possibly involved in deterioration of pancreatic β-cell function and insulin resistance found in type 2 diabetes. Suppression of oxidative stress or the JNK pathway in obese type 2 diabetic mice restored β-cell function and insulin sensitivity, leading to amelioration of glucose tolerance. In addition, oxidative stress and subsequent activation of the JNK pathway are involved in the progression of atherosclerosis, which is often observed as a macroangiopathy under diabetic conditions. Taken together, oxidative stress and subsequent activation of the JNK pathway are closely associated with the pathogenesis of diabetes.

ABBREVIATIONS

Ad, adenovirus; DN, dominant-negative; IRS-1, insulin receptor substrate 1; JNK, c-Jun N-terminal kinase; PDX-1, pancreatic and duodenal homeobox factor-1; ROS, reactive oxygen species; VSMCs, vascular smooth muscle cells; WT, wild type.

REFERENCES

- Aguirre V, Davis R, and White MF. The c-Jun NH₂-terminal kinase promotes insulin resistance during association with insulin receptor substrate-1 and phosphorylation of Ser³⁰⁷. *J Biol Chem* 275: 9047–9054, 2000.
- Ahlgren U, Jonsson J, Jonsson L, Simu K, and Edlund H. β-cell-specific inactivation of the mouse *Ipf1/Pdx1* gene results in loss of the β-cell phenotype and maturity onset diabetes. *Genes Dev* 12: 1763–1768, 1998.
- Ammendrup A, Maillard A, Nielsen K, Aabenhus AN, Serup P, Dragsbaek MO, Mandrup-Poulsen T, and Bonny C. The c-Jun amino-terminal kinase pathway is preferentially activated by interleukin-1 and controls apoptosis in differentiating pancreatic beta-cells. *Diabetes* 49: 1468–1476, 2000
- Andersen HU, Jorgensen KH, Egeberg J, Mandrup-Poulsen T, and Nerup J. Nicotinamide prevents interleukin-1 effects on accumulated insulin release and nitric oxide production in rat islets of Langerhans. *Diabetes* 43: 770–777, 1994.
- Andersen NA, Larsen CM, and Mandrup-Poulsen T. TNFalpha and IFNgamma potentiate IL-1beta induced mitogen activated protein kinase activity in rat pancreatic islets of Langerhans. *Diabetologia* 43: 1389–1396, 2000.
- Ankarcrona M, Dypbukt JM, Brune B, and Nicotera P. Interleukin-1β-induced nitric oxide production activates apoptosis in pancreatic RINm5F cells. Exp Cell Res 213: 172–177, 1994.
- Azumi H, Inoue N, Ohashi Y, Terashima M, Mori T, Fujita H, Awano K, Kobayashi K, Maeda K, Hata K, Shinke T, Kobayashi S, Hirata K, Kawashima S, Itabe H, Hayashi Y, Imajoh-Ohmi S,

- Itoh H, and Yokoyama M. Superoxide generation in directional coronary atherectomy specimens of patients with angina pectoris: important role of NAD(P)H oxidase. *Arterioscler Thromb Vasc Biol* 22: 1838–1844, 2002.
- Barry-Lane P, Patterson C, van der Merwe M, Hu Z, Holland S, Yeh E, and Runge M. p47phox is required for atherosclerotic lesion progression in ApoE(-/-) mice. *J Clin Invest* 108: 1513– 1522, 2001.
- Baynes JW and Thorpe SR. Role of oxidative stress in diabetic complications: a new perspective on an old paradigm. *Diabetes* 48: 1–9, 1999.
- Bertera S, Crawford ML, Alexander AM, Papworth GD, Watkins SC, Robbins PD, and Trucco M. Gene transfer of manganese superoxide dismutase extends islet graft function in a mouse model of autoimmune diabetes. *Diabetes* 52: 387–393, 2003.
- Bonny C, Nicod P, and Waeber G. IB1, a JIP-1-related nuclear protein present in insulin-secreting cells. J Biol Chem 273: 1843–1846, 1998.
- 12. Bonny C, Oberson A, Steinmann M, Schorderet DF, Nicod P, and Waeber G. IB1 reduces cytokine-induced apoptosis of insulin-secreting cells. *J Biol Chem* 275: 16466–16472, 2000.
- Bonny C, Oberson A, Negri S, Sauser C, Schorderet DF. Cell-permeable peptide inhibitors of JNK: novel blockers of beta-cell death. *Diabetes* 50: 77–82, 2001
- Cai H and Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 87: 840–844, 2000.
- Calver A, Collier J, and Vallance P. Inhibition and stimulation of nitric oxide synthesis in the human forearm arterial bed of patients with insulin-dependent diabetes. *J Clin Invest* 90: 2548–2554, 1992
- Chakravarthy U, Hayes RG, Stitt AW, McAuley E, and Archer DB. Constitutive nitric oxide synthase expression in retinal vascular endothelial cells is suppressed by high glucose and advanced glycation end products. *Diabetes* 47: 945–952, 1998.
- Chen H, Li X, and Epstein PN. MnSOD and catalase transgenes demonstrate that protection of islets from oxidative stress does not alter cytokine toxicity. *Diabetes* 54: 1437–1446, 2005.
- Corbett JA, Lancaster JR Jr, Sweetland MA, and McDaniel ML. Interleukin-1β-induced formation of EPR-detectable iron-nitrosyl complexes in islets of Langerhans. *J Biol Chem* 266: 21351– 21354, 1991.
- Corbett JA, Wang JL, Sweetland MA, Jack R, Lancaster JR Jr, and McDaniel ML. Interleukin 1β induces the formation of nitric oxide by β-cells purified from rodent islets of Langerhans. *J Clin Invest* 90: 2384–2391, 1992.
- Corbett JA, and McDaniel ML. Intraislet release of interleukin 1 inhibits beta cell function by inducing beta cell expression of inducible nitric oxide synthase. *J Exp Med* 181: 559–568, 1995.
- Dandona P, Thusu K, Cook S, Snyder B, Makowski J, Armstrong D, and Nicotera T. Oxidative damage to DNA in diabetes mellitus. *Lancet* 347: 444–445, 1996.
- De Keulenaer GW, Alexander RW, Ushio-Fukai M, Ishizaka N, and Griendling KK. Tumour necrosis factor alpha activates a p22phox-based NADH oxidase in vascular smooth muscle. *Biochem J* 329: 653–657, 1998.
- 23. Delafontaine P and Ku L. Reactive oxygen species stimulate insulin-like growth factor I synthesis in vascular smooth muscle cells. *Cardiovasc Res* 33: 216–222, 1997.
- Dickens M, Rogers JS, Cavanagh J, Raitano A, Xia Z, Halpern JR, Greenberg ME, Sawyers CL, and Davis RJ. A cytoplasmic inhibitor of the JNK signal transduction pathway. Science 277: 696–696, 1997.
- Dilts SM, Solvason N, and Lafferty KJ. The role of CD4 and CD8 T cells in the development of autoimmune diabetes. *J Autoimmun* 13: 285–290, 1999.
- Donath MY, Storing J, Maedler K, and Mandrup-Poulsen T. Inflammatory mediators and islet β-cell failure; a link between type 1 and type 2 diabetes. J Mol Med 81: 455–470, 2003.
- Eizirik DL, Pipeleers DG, Ling Z, Welsh N, Hellerstrom C, and Andersson A. Major species differences between humans and ro-

- dents in the susceptibility to pancreatic beta-cell injury. *Proc Natl Acad Sci U S A* 91: 9253–9256, 1994.
- Eizirik DL and Mandrup-Poulsen T. A choice of death: the signaltransduction of immune-mediated beta-cell apoptosis. *Diabetolo*gia 44: 2115–2133, 2001.
- Elliott G and O'Hare P. Intracellular trafficking and protein delivery by a herpesvirus structure protein. Cell 88: 223–233, 1997.
- 30. Evans JL, Goldfine ID, Maddux BA, Grodsky GM. Oxidative stress and stress-activated signaling pathways: a unifying hypothesis of type 2 diabetes. *Endocrine Rev* 23: 599–622, 2002.
- 31. Evans JL, Goldfine ID, Maddux BA, and Grodsky GM. Are oxidative stress-activated signaling pathways mediators of insulin resistance and β-cell dysfunction? *Diabetes* 52: 1–8, 2003.
- 32. Ferber S, Halkin A, Cohen H, Ber I, Einav Y, Goldberg I, Barshack I, Seijffers R, Kopolovic J, Kaiser N, and Karasik A. Pancreatic and duodenal homeobox gene 1 induces expression of insulin genes in liver and ameliorates streptozotocin-induced hyperglycemia. *Nat Med* 6: 568–572, 2000.
- Fortuno A, San Jose G, Moreno MU, Beloqui O, Diez J, and Zalba G. Phagocytic NADPH oxidase overactivity underlies oxidative stress in metabolic syndrome. *Diabetes* 55: 209–215, 2006.
- 34. Frankel AD and Pabo CO. Cellular uptake of the tat protein from human immunodeficiency virus. *Cell* 55: 1189–1193, 1988.
- Gorogawa S, Kajimoto Y, Umayahara U, Kaneto H, Watada H, Kuroda A, Kawamori D, Yasuda T, Matsuhisa M, Yamasaki Y, and Hori M. Probucol preserves pancreatic β-cell function through reduction of oxidative stress in type 2 diabetes. *Diabetes Res Clin Pract* 57: 1–10, 2002.
- Guzik TJ, West NE, Black E, McDonald D, Ratnatunga C, Pillai R, and Channon KM. Functional effect of the C242T polymorphism in the NAD(P)H oxidase p22phox gene on vascular superoxide production in atherosclerosis. *Circulation* 102: 1744–1777, 2000.
- 37. Guzik TJ, Mussa S, Gastaldi D, Sadowski J, Ratnatunga C, Pillai R, Channon KM. Mechanisms of increased vascular superoxide production in human diabetes mellitus: role of NAD(P)H oxidase and endothelial nitric oxide synthase. *Circulation* 105: 1656–1662, 2002.
- Haber CA, Lam TK, Yu Z, Gupta N, Goh T, Bogdanovic E, Giacca A, and Fantus IG. N-acetylcysteine and taurine prevent hyperglycemia-induced insulin resistance in vivo: possible role of oxidative stress. *Am J Physiol Endocrinol Metab* 285: E744–E753, 2003.
- 39. Harmon JS, Stein R, and Robertson RP. Oxidative stress-mediated, post-translational loss of MafA protein as a contributing mechanism to loss of insulin gene expression in glucotoxic beta cells. *J Biol Chem* 280: 11107–11113, 2005.
- Harrison D, Griendling KK, Landmesser U, Hornig B, and Drexler H. Role of oxidative stress in atherosclerosis. *Am J Cardiol* 91: 7A–11A, 2003.
- 41. Hayaishi-Okano R, Yamasaki Y, Kajimoto Y, Sakamoto K, Ohtoshi K, Katakami N, Kawamori D, Miyatsuka K, Hatazaki M, Hazama Y, and Hori M. Association of NAD(P)H oxidase p22 phox gene variation with advanced carotid atherosclerosis in Japanese type 2 diabetes. *Diabetes Care* 262: 458–463, 2003.
- 42. Hink U, Li H, Mollnau H, Oelze M, Matheis E, Hartmann M, Skatchkov M, Thaiss F, Stahl RA, Warnholtz A, Meinertz T, Griendling K, Harrison DG, Forstermann U, and Munzel T. Mechanisms underlying endothelial dysfunction in diabetes mellitus. *Circ Res* 88: E14–E22, 2001.
- Hirosumi J, Tuncman G, Chang L, Karin M, and Hotamisligil GS. A central role for JNK in obesity and insulin resistance. *Nature* 420: 333–336, 2002.
- 44. Hohmeier HE, Thigpen A, Tran VV, Davis R, and Newgard CB. Stable expression of manganese superoxide dismutase (MnSOD) in insulinoma cells prevents IL-1beta-induced cytotoxicity and reduces nitric oxide production. *J Clin Invest* 101: 1811–1820, 1009
- Holland AM, Hale MA, Kagami H, Hammer RE, and MacDonald RJ. Experimental control of pancreatic development and maintenance. *Proc Natl Acad Sci USA* 99: 12236–12241, 2002.

 Hotamisligil G. Role of endoplasmic reticulum stress and c-Jun NH2-terminal kinase pathways in inflammation and origin of obesity and diabetes. *Diabetes* 54: S73–S78, 2005.

- 47. Hotta M, Tashiro F, Ikegami H, Niwa H, Ogihara T, Yodoi J, and Miyazaki J: Pancreatic beta cell-specific expression of thioredoxin, an antioxidative and antiapoptotic protein, prevents autoimmune and streptozotocin-induced diabetes. *J Exp Med* 188: 1445–1451, 1998.
- Hu Y, Cheng L, Hochleitner BW, and Xu Q. Activation of mitogen-activated protein kinases (ERK/JNK) and AP-1 transcription factor in rat carotid arteries after balloon injury. Arterioscler Thromb Vasc Biol 17: 2808–2816, 1997.
- Ihara Y, Toyokuni S, Uchida K, Odaka H, Tanaka T, Ikeda H, Hiai H, Seino Y, and Yamada Y. Hyperglycemia causes oxidative stress in pancreatic β-cells of GK rats, a model of type 2 diabetes. *Diabetes* 48: 927–932, 1999.
- Inoguchi T, Li P, Umeda F, Yu HY, Kakimoto M, Imamura M, Aoki T, Etoh T, Hashimoto T, Naruse M, Sano H, Utsumi H, and Nawata H. High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 49: 1939–1945, 2000.
- 51. Izumi Y, Kim S, Namba M, Yasumoto H, Miyazaki H, Hoshiga M, Kaneda Y, Morishita R, Zhan Y, and Iwao H. Gene transfer of dominant-negative mutants of extracellular signal-regulated kinase and c-Jun NH₂-terminal kinase prevents neointimal formation in balloon-injured rat artery. Circ Res 88: 1120–1126, 2001.
- 52. Jacob S, Ruus P, Hermann R, Tritschler HJ, Maerker E, Renn W, Augustin HJ, Dietze GJ, and Rett K. Oral administration of RAC-alpha-lipoic acid modulates insulin sensitivity in patients with type-2 diabetes mellitus: a placebo-controlled pilot trial. Free Radic Biol Med 27: 309–314, 1999.
- 53. Jaeschke A, Rincon M, Doran B, Reilly J, Neuberg D, Greiner DL, Shultz LD, Rossini AA, Flavell RA, and Davis RJ. Disruption of the Jnk2 (Mapk9) gene reduces destructive insulitis and diabetes in a mouse model of type I diabetes. *Proc Natl Acad Sci U S A* 102: 6931–6935, 2005.
- 54. Jonas JC, Sharma A, Hasenkamp W, Iikova H, Patane G, Laybutt R, Bonner-Weir S, and Weir GC. Chronic hyperglycemia triggers loss of pancreatic β cell differentiation in an animal model of diabetes. *J Biol Chem* 274: 14112–14121, 1999.
- Jonsson J, Carlsson L, Edlund T, and Edlund H. Insulin-promoterfactor 1 is required for pancreas development in mice. *Nature* 37: 606–609, 1994.
- Kajimoto Y, Matsuoka T, Kaneto H, Watada H, Fujitani Y, Kishimoto M, Sakamoto K, Matsuhisa M, Kawamori R, Yamasaki Y, and Hori M. Induction of glycation suppresses glucokinase gene expression in HIT-T15 cells. *Diabetologia* 42: 1417–1424, 1999.
- Kaneto H, Fujii J, Seo HG, Suzuki K, Matsuoka T, Nakamura N, Tatsumi H, Yamasaki Y, Kamada T, and Taniguchi N. Apoptotic cell death triggered by nitric oxide in pancreatic β-cells. *Diabetes* 44: 733–738, 1995.
- 58. Kaneto H, Kajimoto Y, Miyagawa J, Matsuoka T, Fujitani Y, Umayahara Y, Hanafusa T, Matsuzawa Y, Yamasaki Y, and Hori M. Beneficial effects of antioxidants for diabetes: possible protection of pancreatic β-cells against glucose toxicity. *Diabetes* 48: 2398–2406, 1999.
- Kaneto H, Xu G, Song KH, Suzuma K, Bonner-Weir S, Sharma A, and Weir GC. Activation of the hexosamine pathway leads to deterioration of pancreatic β-cell function by provoking oxidative stress. *J Biol Chem* 276: 31099–31104, 2001.
- Kaneto H, Xu G, Fujii N, Kim S, Bonner-Weir S, and Weir GC. Involvement of c-Jun N-terminal kinase in oxidative stress-mediated suppression of insulin gene expression. *J Biol Chem* 277: 30010–30018, 2002.
- Kaneto H, Nakatani Y, Miyatsuka T, Kawamori D, Matsuoka T, Matsuhisa M, Kajimoto Y, Ichijo H, Yamasaki Y, and Hori M. Possible novel therapy for diabetes with cell-permeable JNK inhibitory peptide. *Nat Med* 10: 1128–1132, 2004.
- Kaneto H, Matsuoka T, Nakatani Y, Kawamori D, Miyatsuka T, Matsuhisa M, and Yamasaki Y. Oxidative stress, ER stress, and the JNK pathway in type 2 diabetes. J Mol Med 83: 429–439, 2005.

 Kaneto H, Matsuoka T, Nakatani Y, Miyatsuka T, Matsuhisa M, Hori M, and Yamasaki Y. A crucial role of MafA as a novel therapeutic target for diabetes. *J Biol Chem* 280: 15047–15052, 2005.

- 64. Katakami N, Kaneto H, Hao H, Umayahara Y, Fujitani Y, Sakamoto K, Gorogawa S, Yasuda T, Kawamori D, Kajimoto Y, Matsuhisa M, Yutani C, Hori M, and Yamasaki Y. Role of pim-1 in smooth muscle cell proliferation. *J Biol Chem* 279: 54742–54749, 2004
- Katakami N, Yamasaki Y, Hayaishi R, Ohtoshi K, Kaneto H, Matsuhisa M, Kosugi K, and Hori M. Metformin or gliclazide, rather than glibenclamide, attenuate progression of carotid intima-media thickness in subjects with type 2 diabetes. *Diabetologia* 47: 1906–1923, 2004.
- 66. Kataoka K, Han SI, Shioda S, Hirai M, Nishizawa M, Handa H. MafA is a glucose-regulated and pancreatic β-cell-specific transcriptional activator for the insulin gene. *J Biol Chem* 277: 49903–49910, 2002.
- 67. Kawamori D, Kajimoto Y, Kaneto H, Umayahara Y, Fujitani Y, Miyatsuka T, Watada H, Leibiger IB, Yamasaki Y, and Hori M. Oxidative stress induces nucleo-cytoplasmic translocation of pancreatic transcription factor PDX-1 through activation of c-Jun N-terminal kinase. *Diabetes* 52: 2896–2904, 2003.
- 68. Kawamori D, Kaneto H, Nakatani Y, Matsuoka T, Matsuhisa M, Hori M, and Yamasaki Y. The forkhead transcription factor Foxol bridges the JNK pathway and the transcription factor PDX-1 through its intracellular translocation. *J Biol Chem* 281: 1091–1098, 2006.
- Kawamori R, Yamasaki Y, Matsushima H, Nishizawa H, Nao K, Hougaku H, Maeda H, Handa N, Matsumoto M, and Kadada T. Prevalence of carotid atherosclerosis in diabetic patients: ultrasound high-resolution B-mode imaging on carotid arteries. *Diabetes Care* 15: 1290–1294, 1992.
- Kim S, Izumi Y, Yano M, Hamaguchi A, Miura K, Yamanaka S, Miyazaki H, and Iwao H. Angiotensin blockade inhibits activation of mitogen-activated protein kinases in rat balloon-injured artery. Circulation 97: 1731–1737, 1998.
- Kim YK, Lee MS, Son SM, Kim IJ, Lee WS, Rhim BY, Hong KW, and Kim CD. Vascular NADH oxidase is involved in impaired endothelium-dependent vasodilation in OLETF rats, a model of type 2 diabetes. *Diabetes* 51: 522–527, 2002.
- 72. Konrad T, Vicini P, Kusterer K, Hoflich A, Assadkhani A, Bohles HJ, Sewell A, Tritschler HJ, Cobelli C, and Usadel KH. a-Lipoic acid treatment decreases serum lactate and pyruvate concentrations and improves glucose effectiveness in lean and obese patients with type 2 diabetes. *Diabetes Care* 22: 280–287, 1999.
- Koyama H, Olson NE, Dastvan FF, and Reidy MA. Cell replication in the arterial wall: activation of signaling pathway following in vivo injury. Circ Res 82: 713–721, 1998.
- Kubisch HM, Wang J, Luche R, Carlson E, Bray TM, Epstein CJ, and Phillips JP. Transgenic copper/zinc superoxide dismutase modulates susceptibility to type I diabetes. *Proc Natl Acad Sci U S* A 91: 9956–9959, 1994.
- Kubisch HM, Wang JQ, Bray TM, and Phillips JP. Targeted overexpression of Cu/Zu superoxide-dismutase protects pancreatic beta-cells against oxidative stress. *Diabetes* 46: 1563–1566, 1997.
- Larsen CM, Wadt KA, Juhl LF, Andersen HU, Karlsen AE, Su MS, Seedorf K, Shapiro L, Dinarello CA, and Mandrup-Poulsen T. Interleukin-1beta-induced rat pancreatic islet nitric oxide synthesis requires both the p38 and extracellular signal-regulated kinase 1/2 mitogen-activated protein kinases. *J Biol Chem* 273: 152940–153001, 1998.
- 77. Lehtinen MK, Yuan Z, Boag PR, Yang Y, Villen J, Becker EBE, DiBacco S, de la Iglesia N, Gygi S, Blackwell TK, and Bonni A. A conserved MST-FOXO signaling pathway mediates oxidative-stress responses and extends life span. *Cell* 125: 987–1001, 2006.
- Lenzen S, Drinkgern J, and Tiedge M. Low antioxidant enzyme gene expression in pancreatic islets compared with various other mouse tissues. Free Radic Biol Med 20: 463

 –466, 1996.
- Leonard J, Peers B, Johnson T, Ferreri K, Lee S, and Montminy MR. Characterization of somatostatin transactivating factor-1, a novel homeobox factor that stimulates somatostatin expression in pancreatic islet cells. *Mol Endocrinol* 7: 1275–1283, 1993.

- Lortz S, Tiedge M, Nachtwey T, Karlsen AE, Nerup J, and Lenzen S. Protection of insulin-producing RINm5F cells against cytokine-mediated toxicity through overexpression of antioxidant enzymes. *Diabetes* 49: 1123–1130, 2000.
- 81. Madamanchi NR, Vendrov A, and Runge MS. Oxidative stress and vascular disease. *Artheroscler Thromb Vasc Biol* 25: 29–35, 2005.
- Maechler P, Jornot L, and Wollheim CB. Hydrogen peroxide alters mitochondrial activation and insulin secretion in pancreatic beta cells. *J Biol Chem* 274: 27905–27913, 1999
- Major CD and Wolf BA. Interleukin-1beta stimulation of c-Jun NH (2)-terminal kinase activity in insulin-secreting cells: evidence for cytoplasmic restriction. *Diabetes* 50: 2721–2728, 2001.
- Mandrup-Poulsen T. Beta cell death and protection. Ann NY Acad Sci 1005: 32–42, 2003.
- Matsuoka T, Kajimoto Y, Watada H, Kaneto H, Kishimoto M, Umayahara Y, Fujitani Y, Kamada T, Kawamori R, and Yamasaki Y. Glycation-dependent, reactive oxygen species-mediated suppression of the insulin gene promoter activity in HIT cells. *J Clin Invest* 99: 144–150, 1997
- Matsuoka T, Zhao L, Artner I, Jarrett HW, Friedman D, Means A, and Stein R. Members of the large Maf transcription family regulate insulin gene transcription in islet β cells. *Mol Cell Biol* 23: 6049–6062, 2003.
- 87. Matsuoka T, Artner I, Henderson E, Means A, Sander M, and Stein R. The MafA transcription factor appears to be responsible for tissue-specific expression of insulin. *Proc Natl Acad Sci U S A* 101: 2930–2933, 2004.
- Mehta JL, Rasouli N, Sinha AK, and Molavi B. Mehta JL, Rasouli N, Sinha AK Oxidative stress in diabetes: a mechanistic overview of its effects on atherogenesis and myocardial dysfunction. *Int J Biochem Cell Biol* 38: 794–803, 2006.
- Miller CP, McGehee RE, and Habener JF. IDX-1: a new homeodomain transcription factor expressed in rat pancreatic islets and duodenum that transactivates the somatostatin gene. *EMBO J* 13: 1145–1156, 1994.
- Miyazaki S, Yamato E, and Miyazaki J. Regulated expression of pdx-1 promotes in vitro differentiation of insulin-producing cells from embryonic stem cells. *Diabetes* 53: 1030–1037, 2004.
- Miyazaki S, Miyazaki T, Tashiro F, Yamato E, and Miyazaki, J. Development of a single-cassette system for spatiotemporal gene regulation in mice. *Biochem Biophys Res Commun* 338, 1083– 1088, 2005
- Mohazzab KM, Kaminski PM, and Wolin MS. NADH oxidoreductase is a major source of superoxide anion in bovine coronary artery endothelium. *Am J Phyiol* 266: H2568–H2572, 1994.
- 93. Moran A, Zhang H-J, Olson LK, Harmon JS, Poitout V, and Robertson RP. Differentiation of glucose toxicity from beta cell exhaustion during the evolution of defective insulin gene expression in the pancreatic islet cell line, HIT-T15. *J Clin Invest* 99: 534–539, 1997.
- Moritoh Y, Yamato E, Yasui Y, Miyazaki S, and Miyazaki J. Analysis of insulin-producing cells during in vitro differentiation from feeder-free embryonic stem cells. *Diabetes* 52: 1163–1168, 2003.
- Nagahara H, Vocero-Akbani AM, Snyder EL, Ho A, Latham DG, Lissy NA, Becker-Hapak M, Ezhevsky SA, and Dowdy SF. Transduction of full-length TAT fusion proteins into mammalian cells: TAT-p-27^{Kip1} induces cell migration. *Nat Med* 4: 1449–1452, 1998.
- Nakatani Y, Kaneto H, Kawamori D, Hatazaki M, Miyatsuka T, Matsuoka T, Kajimoto Y, Matsuhisa M, Yamasaki Y, and Hori M. Modulation of the JNK pathway in liver affects insulin resistance status. J Biol Chem 279: 45803–45809, 2004.
- Noguchi H, Kaneto H, Weir GC, Bonner-Weir S. PDX-1 protein containing its own Antennapedia-like protein transduction domain can transduce pancreatic duct and islet cells. *Diabetes* 52: 1732–1737, 2003.
- Noguchi H, Matsushita M, Okitsu T, Moriwaki A, Tomizawa K, Kang S, Li ST, Kobayashi N, Matsumoto S, Tanaka K, Tanaka N, and Matsui H. A new cell-permeable peptide allows successful allogeneic islet transplantation in mice. *Nat Med* 10: 305–309, 2004
- Ohlsson H, Karlsson K, and Edlund T. IPF1, a homeodomaincontaining-transactivator of the insulin gene. *EMBO J* 12: 4251–4259, 1993.

- Olbrot M, Rud J, Moss LG, and Sharma A. Identification of βcell-specific insulin gene transcription factor RIPE3b1 as mammalian MafA. Proc Natl Acad Sci U S A 99: 6737–6742, 2002.
- 101. Patterson C, Ruef J, Madamanchi NR, Barry-Lane P, Hu Z, Horaist C, Ballinger CA, Brasier AR, Bode C, and Runge MS. Stimulation of a vascular smooth muscle cell NAD(P)H oxidase by thrombin: evidence that p47(phox) may participate in forming this oxidase in vitro and in vivo. *J Biol Chem* 274: 19814–19822, 1999.
- 102. Piganelli JD, Flores SC, Cruz C, Koepp J, Batinic-Haberle I, Crapo J, Day B, Kachadourian R, Young R, Bradley B, and Haskins K. A metalloporphyrin-based superoxide dismutase mimic inhibits adoptive transfer of autoimmune diabetes by a diabetogenic T-cell clone. *Diabetes* 51: 347–355, 2002.
- 103. Poitout V, Olson LK, and Robertson RP. Chronic exposure of bTC-6 cells to supraphysiologic concentrations of glucose decreases binding of the RIPE3b1 insulin gene transcription activator. J Clin Invest 97: 1041–1046, 1996.
- 104. Poitout V and Robertson RP. Minireview: secondary beta-cell failure in type 2 diabetes: a convergence of glucotoxicity and lipotoxicity. *Endocrinology* 143: 339–342, 2002.
- 105. Rabinovitch A. Immunoregulatory and cytokine imbalances in the pathogenesis of IDDM: therapeutic intervention by immunostimulation? *Diabetes* 43: 613–621, 1994.
- Rao GN and Berk BC. Active oxygen species stimulate vascular smooth muscle cell growth and proto-oncogene expression. *Circ Res* 70: 593–599, 1992.
- 107. Ricci R, Sumara G, Sumara I, Rozenberg I, Kurrer M, Akhmedov A, Hersberger M, Eriksson U, Eberli FR, Becher B, Boren J, Chen M, Cybulsky MI, Moore KJ, Freeman MW, Wagner EF, Matter CM, and Luscher TF. Requirement of JNK2 for scavenger receptor A-mediated foam cell formation in atherogenesis. *Science* 306: 1558–1561, 2004.
- 108. Robertson RP, Harmon J, Tran PO, Tanaka Y, and Takahashi H. Glucose toxicity in β-cells: type 2 diabetes, good radicals gone bad, and the glutathione connection. *Diabetes* 52: 581–587, 2003
- Robertson RP. Chronic oxidative stress as a central mechanism for glucose toxicity in pancreatic islet beta cells in diabetes. J Biol Chem 271: 42351–42354, 2004
- Rothbard JB, Garlington S, Lin Q, Kirschberg T, Kreider E, Mc-Grane PL, Wender PA, and Khavari PA. Conjugation of arginine oligomers to cyclosporin A facilitates topical delivery and inhibition of inflammation. *Nat Med* 6: 1253–1257, 2000.
- 111. Rudich A, Tirosh A., Potashnik R, Hemi R, Kanety H, and Bashan N. Prolonged oxidative stress impairs insulin-induced GLUT4 translocation in 3T3-L1 adipocytes. *Diabetes* 47: 1562–1569, 1998.
- 112. Sakai K, Matsumoto K, Nishikawa T, Suefuji M, Nakamaru K, Hirashima Y, Kawashima J, Shirotani T, Ichinose K, Brownlee M, and Araki E. Mitochondrial reactive oxygen species reduce insulin secretion by pancreatic β-cells. *Biochem Biophys Res Commun* 300: 216–222, 2003.
- 113. Sakamoto K, Yamasaki Y, Kaneto H, Fujitani Y, Matsuoka T, Yoshioka R, Tagawa T, Kajimoto Y, and Hori M. Identification of oxidative stress-regulated genes in rat aortic smooth muscle cells by suppression subtractive hybridization. FEBS Lett 12: 47–51, 1999
- Schwarze SR, Ho A, Vocero-Akbani AM, and Dowdy SF. In vivo protein transduction: delivery of a biologically active protein into the mouse. *Science* 285: 1569–1572, 1999.
- 115. Sharma A, Fusco-DeMane D, Henderson E, Efrat S, and Stein R. The role of the insulin control element and RIPE3b1 activators in glucose-stimulated transcription of the insulin gene. *Mol Endocrinol* 9: 1468–1488, 1995.
- Shimabukuro M, Ohneda M, Lee Y, and Unger RH. Role of nitric oxide in obesity-induced beta cell disease. *J Clin Invest* 46: 1276–1280, 1997.
- Shimabukuro M, Zhou YT, Levi M, and Unger RH. Fatty acidinduced beta cell apoptosis: a link between obesity and diabetes. *Proc Natl Acad Sci U S A* 95: 2498–2502, 1998.
- 118. Stephens NG, Parsons A, Schofield PM, Kelly F, Cheeseman K, and Mitchinson MJ. Randomised controlled trial of vitamin E in

patients with coronary disease: Cambridge Heart Antioxidant Study (CHAOS). *Lancet* 347: 781–786, 1996.

- Stoffers DA, Zinkin NT, Stanojevic V, Clarke WL, and Habener JF.
 Pancreatic agenesis attributable to a single nucleotide deletion in the human IPF1 gene coding sequence. Nat Genet 15: 106–110, 1997.
- Suarez-Pinzon WL, Szabo C, and Rabinovitch A. Development of autoimmune diabetes in NOD mice is associated with the formation of peroxynitrite in pancreatic islet beta-cells. *Diabetes* 46: 907–911, 1997.
- Sundaresan M, Yu ZX, Ferrans VJ, Irani K, and Finkel T. Requirement for generation of H₂O₂ for platelet-derived growth factor signal transduction. *Science* 270: 296–299, 1995.
- 122. Tabatabaie T, Vasquez-Weldon A, Moore DR, and Kotake Y. Free radicals and the pathogenesis of type 1 diabetes: β-cell cytokinemediated free radical generation via cyclooxygenase-2. *Diabetes* 52: 1994–1999, 2003.
- Tanaka Y, Gleason CE, Tran POT, Harmon JS, and Robertson RP. Prevention of glucose toxicity in HIT-T15 cells and Zucker diabetic fatty rats by antioxidants. *Proc Natl Acad Sci U S A* 96: 10857–10862, 1999.
- 124. Tanaka Y, Tran POT, Harmon J, and Robertson RP. A role of glutathione peroxidase in protecting pancreatic β cells against oxidative stress in a model of glucose toxicity. *Proc Natl Acad Sci U S A* 99: 12363–12368, 2002.
- 125. Taniguchi H, Yamato E, Tashiro F, Ikegami H, Ogihara T, and Miyazaki J. β-Cell neogenesis induced by adenovirus-mediated gene delivery of transcription factor pdx-1 into mouse pancreas. Gene Ther 10: 15–23, 2003.
- 126. Tardif JC, Cote G, Lesperance J, Bourassa M, Lambert J, Doucet S, Bilodeau L, Nattel S, and de Guise P. Probucol and multivitamins in the prevention of restenosis after coronary angioplasty: Multivitamins and Probucol Study Group. N Engl J Med 337: 365–372, 1997.
- 127. Tiedge M, Lortz S, Munday R, and Lenzen S. Complementary action of antioxidant enzymes in the protection of bioengineered insulin-producing RINm5F cells against the toxicity of reactive oxygen species. *Diabetes* 47: 1578–1585, 1998.
- 128. Tirosh A, Potashnik R, Bashan N, and Rudich A. Oxidative stress disrupts insulin-induced cellular redistribution of insulin receptor substrate-1 and phosphatidylinositol 3-kinase in 3T3-L1 adipocytes: a putative cellular mechanism for impaired protein kinase B activation and GLUT4 translocation. *J Biol Chem* 274: 10595–10602, 1999.
- 129. Ushio-Fukai M, Zafari AM, Fukui T, Ishizaka N, and Griendling KK. p22phox is a critical component of the superoxide-generating NADH/NADPH oxidase system and regulates angiotensin II-induced hypertrophy in vascular smooth muscle cells. *J Biol Chem* 271: 23317–23321, 1996.
- 130. Waeber G, Delplanque J, Bonny C, Mooser V, Steinmann M, Widmann C, Maillard A, Miklossy J, Dina C, Hani EH, Vionnet N, Nicod P, Boutin P, and Froguel P. The gene MAPK8IP1, en-

- coding islet-brain-1, is a candidate for type 2 diabetes. *Nat Genet* 24: 291–295, 2000.
- Weir GC, Laybutt DR, Kaneto H, Bonner-Weir S, and Sharma A. β-Cell adaptation and decompensation during the progression of diabetes. *Diabetes* 50: S154–S159, 2001.
- 132. Wellen KE and Hotamisligil G. Inflammation, stress, and diabetes. *J Clin Invest* 115: 1111–1119, 2005.
- 133. Welsh N. Interleukin-1 beta-induced ceramide and diacylglycerol generation may lead to activation of the c-Jun NH2-terminal kinase and the transcription factor ATF2 in the insulin-producing cell line RINm5F. *J Biol Chem* 271: 8307–8312, 1996.
- 134. Wong FS and Janeway CA Jr. The role of CD4 vs. CD8 T cells in IDDM. *J Autoimmun* 13: 290–295, 1999.
- 135. Yamasaki Y, Kawamori R, Matsushima H, Nishizawa H, Kodama M, Kajimoto Y, Morishima T, and Kadada T. Atherosclerosis in carotid artery of young IDDM patients monitored by ultrasound high-resolution B-mode imaging. *Diabetes* 43: 634–639, 1994.
- 136. Yamasaki Y, Kawamori R, Matsushima H, Nishizawa H, Kodama M, Kubota M, Kajimoto Y, and Kadada T. Asymptomatic hyperglycaemia is associated with increased intimal plus medial thickness of the carotid artery. *Diabetologia* 38: 585–591, 1995.
- 137. Yamasaki Y, Katakami N, Hayaishi-Okano R, Matsuhisa M, Kajimoto Y, Kosugi K, Hatano M, and Hori M. a-Glucosidase inhibitor reduces the progression of carotid intima-media thickness. *Diabetes Res Clin Pract* 67: 204–210, 2005.
- 138. Yoshizumi M, Abe J, Haendeler J, Huang Q, and Berk BC. Src and Cas mediate JNK activation but not ERK1/2 and p38 kinases by reactive oxygen species. *J Biol Chem* 275: 11706–11712, 2000
- 139. Zhang C, Moriguchi T, Kajihara M, Esaki R, Harada A, Shimohata H, Oishi H, Hamada M, Morito N, Hasegawa K, Kudo T, Engel JD, Yamamoto M, and Takahashi S. MafA is a key regulator of glucose-stimulated insulin secretion. *Mol Cell Biol* 25: 4969–4976, 2005.

Address correspondence to: Hideaki Kaneto, MD, PhD Department of Internal Medicine and Therapeutics Osaka University Graduate School of Medicine 2–2 Yamadaoka, Suita Osaka 565–0871, Japan

E-mail: kaneto@medone.med.osaka-u.ac.jp

Date of first submission to ARS Central, September 20, 2006; date of acceptance, September 26, 2006.

This article has been cited by:

- 1. A. T. Treweeke, T. J. Winterburn, I. Mackenzie, F. Barrett, C. Barr, G. F. Rushworth, I. Dransfield, S. M. MacRury, I. L. Megson. 2012. N-Acetylcysteine inhibits platelet—monocyte conjugation in patients with type 2 diabetes with depleted intraplatelet glutathione: a randomised controlled trial. *Diabetologia* 55:11, 2920-2928. [CrossRef]
- 2. T R Neyestani, Z Ghandchi, M-R Eshraghian, A Kalayi, N Shariatzadeh, A Houshiarrad. 2012. Evidence for augmented oxidative stress in the subjects with type 1 diabetes and their siblings: a possible preventive role for antioxidants. *European Journal of Clinical Nutrition* 66:9, 1054-1058. [CrossRef]
- 3. N. Poungvarin, J. K. Lee, V. K. Yechoor, M. V. Li, T. Assavapokee, P. Suksaranjit, J. J. Thepsongwajja, P. K. Saha, K. Oka, L. Chan. 2012. Carbohydrate response element-binding protein (ChREBP) plays a pivotal role in beta cell glucotoxicity. *Diabetologia*. [CrossRef]
- 4. Muhammad Sajid Hamid Akash, Qi Shen, Kanwal Rehman, Shuqing Chen. 2012. Interleukin-1 receptor antagonist: A new therapy for type 2 diabetes mellitus. *Journal of Pharmaceutical Sciences* n/a-n/a. [CrossRef]
- 5. Karthikeyan Chandrasekaran, Kavitha Swaminathan, S. Mathan Kumar, Dahn L. Clemens, Aparajita Dey. 2012. Increased oxidative stress and toxicity in ADH and CYP2E1 overexpressing human hepatoma VL-17A cells exposed to high glucose. *Integrative Biology*. [CrossRef]
- 6. Maher Boukhris, Mohamed Bouaziz, Ines Feki, Hedya Jemai, Abdelfattah El Feki, Sami Sayadi. 2012. Hypoglycemic and antioxidant effects of leaf essential oil of Pelargonium graveolens L'Hér. in alloxan induced diabetic rats. *Lipids in Health and Disease* 11:1, 81. [CrossRef]
- 7. Marc Prentki, S.R. Murthy Madiraju. 2011. Glycerolipid/free fatty acid cycle and islet #-cell function in health, obesity and diabetes. *Molecular and Cellular Endocrinology*. [CrossRef]
- 8. F. Bobeuf, M. Labonte, I. J. Dionne, Abdelouahed Khalil. 2011. Combined effect of antioxidant supplementation and resistance training on oxidative stress markers, muscle and body composition in an elderly population. *The journal of nutrition, health & aging*. [CrossRef]
- 9. Chun Fa Huang, Ya Wen Chen, Ching Yao Yang, Keh Sung Tsai, Rong Sen Yang, Shing Hwa Liu. 2011. Arsenic and diabetes: Current perspectives. *The Kaohsiung Journal of Medical Sciences*. [CrossRef]
- 10. Junling Yang, Qianfei Xue, Lining Miao, Lu Cai. 2011. Pulmonary fibrosis: a possible diabetic complication. *Diabetes/Metabolism Research and Reviews* 27:4, 311-317. [CrossRef]
- 11. Daolin Tang, Rui Kang, Herbert J. Zeh III, Michael T. Lotze. 2011. High-Mobility Group Box 1, Oxidative Stress, and Disease. *Antioxidants & Redox Signaling* 14:7, 1315-1335. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 12. Hye Duck Choi, Ji Hae Kim, Min Jung Chang, Yeo Kyu-Youn, Wan Gyoon Shin. 2011. Effects of Astaxanthin on Oxidative Stress in Overweight and Obese Adults. *Phytotherapy Research* n/a-n/a. [CrossRef]
- 13. Deirdre C. Keane, Hilton K. Takahashi, Shalinee Dhayal, Noel G. Morgan, Rui Curi, Philip Newsholme. 2011. Arachidonic acid actions on functional integrity and attenuation of the negative effects of palmitic acid in a clonal pancreatic #-cell line. *Clinical Science* 120:5, 195-206. [CrossRef]
- 14. Chang Yeob Han, Sung Hwan Ki, Young Woo Kim, Kyoung Noh, Da Yeon Lee, Bomi Kang, Jae-Ha Ryu, Raok Jeon, Eun Hyun Kim, Se Jin Hwang, Sang Geon Kim. 2011. Ajoene, A Stable Garlic By-Product, Inhibits High Fat Diet-Induced Hepatic Steatosis and Oxidative Injury Through LKB1-Dependent AMPK Activation. *Antioxidants & Redox Signaling* 14:2, 187-202. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF] with Links]
- 15. Aboozar Monavar Feshani, Shideh Monatasser Kouhsari, Saeed Mohammadi. 2011. Vaccinium arctostaphylos, a common herbal medicine in Iran: Molecular and biochemical study of its antidiabetic effects on alloxan-diabetic Wistar rats. *Journal of Ethnopharmacology* **133**:1, 67-74. [CrossRef]
- 16. Marta Michalska, Gabriele Wolf, Reinhard Walther, Philip Newsholme. 2010. Effects of pharmacological inhibition of NADPH oxidase or iNOS on pro-inflammatory cytokine, palmitic acid or H 2 O 2 -induced mouse islet or clonal pancreatic #-cell dysfunction. *Bioscience Reports* 30:6, 445-453. [CrossRef]
- 17. G. Leibowitz, E. Bachar, M. Shaked, A. Sinai, M. Ketzinel-Gilad, E. Cerasi, N. Kaiser. 2010. Glucose regulation of #-cell stress in type 2 diabetes. *Diabetes, Obesity and Metabolism* 12, 66-75. [CrossRef]
- 18. Guadalupe Sabio, Roger J. Davis. 2010. cJun NH2-terminal kinase 1 (JNK1): roles in metabolic regulation of insulin resistance. *Trends in Biochemical Sciences* **35**:9, 490-496. [CrossRef]

- 19. Muhammad R. Baig, Erica Navaira, Michael A. Escamilla, Henriette Raventos, Consuelo Walss-Bass. 2010. Clozapine Treatment Causes Oxidation of Proteins Involved in Energy Metabolism in Lymphoblastoid Cells: A Possible Mechanism for Antipsychotic- Induced Metabolic Alterations. *Journal of Psychiatric Practice* 16:5, 325-333. [CrossRef]
- 20. Amadou K.S. Camara, Edward J. Lesnefsky, David F. Stowe. 2010. Potential Therapeutic Benefits of Strategies Directed to Mitochondria. *Antioxidants & Redox Signaling* 13:3, 279-347. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF with Links]
- 21. Stefania Cannito, Erica Novo, Lorenzo Valfrè di Bonzo, Chiara Busletta, Sebastiano Colombatto, Maurizio Parola. 2010. Epithelial–Mesenchymal Transition: From Molecular Mechanisms, Redox Regulation to Implications in Human Health and Disease. Antioxidants & Redox Signaling 12:12, 1383-1430. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF] with Links]
- 22. Jyoti D. Malhotra, Randal J. KaufmanEndoplasmic Reticulum Stress and Oxidative Stress: Mechanisms and Link to Disease 21-46. [CrossRef]
- 23. S. Sivabalan, C.V. Anuradha. 2010. A Comparative Study on the Antioxidant and Glucose-lowering Effects of Curcumin and Bisdemethoxycurcumin Analog through in vitro Assays. *International Journal of Pharmacology* **6**:5, 664-669. [CrossRef]
- 24. Karthikeyan Chandrasekaran, Kavitha Swaminathan, Suvro Chatterjee, Aparajita Dey. 2010. Apoptosis in HepG2 cells exposed to high glucose. *Toxicology in Vitro* **24**:2, 387-396. [CrossRef]
- 25. Gyorgy Szabadkai, Michael R. Duchen. 2009. Mitochondria mediated cell death in diabetes. *Apoptosis* **14**:12, 1405-1423. [CrossRef]
- 26. P. Newsholme, D. Morgan, E. Rebelato, H. C. Oliveira-Emilio, J. Procopio, R. Curi, A. Carpinelli. 2009. Insights into the critical role of NADPH oxidase(s) in the normal and dysregulated pancreatic beta cell. *Diabetologia* **52**:12, 2489-2498. [CrossRef]
- 27. Nirit Hanin-Avraham, Bianca Fuhrman, Agnieszka Mech-Dorosz, Sofiya Kolusheva, Angel Porgador, Michael Aviram, Raz Jelinek. 2009. Lipoprotein interactions with chromatic membranes as a novel marker for oxidative stress-related diseases. *Biochimica et Biophysica Acta (BBA) Biomembranes* 1788:11, 2436-2443. [CrossRef]
- 28. José Serrano, Riitta Puupponen-Pimiä, Andreas Dauer, Anna-Marja Aura, Fulgencio Saura-Calixto. 2009. Tannins: Current knowledge of food sources, intake, bioavailability and biological effects. *Molecular Nutrition & Food Research* **53**:S2, S310-S329. [CrossRef]
- 29. Rui Guo, Heng Ma, Feng Gao, Li Zhong, Jun Ren. 2009. Metallothionein alleviates oxidative stress-induced endoplasmic reticulum stress and myocardial dysfunction. *Journal of Molecular and Cellular Cardiology* **47**:2, 228-237. [CrossRef]
- 30. Jasemine Shabeer, Radhey Shyam Srivastava, Sushil Kumar Singh. 2009. Antidiabetic and antioxidant effect of various fractions of Phyllanthus simplex in alloxan diabetic rats. *Journal of Ethnopharmacology* **124**:1, 34-38. [CrossRef]
- 31. Jiejie Hao, Weili Shen, Chuan Tian, Zhongbo Liu, Jinmin Ren, Cheng Luo, Jiangang Long, Edward Sharman, Jiankang Liu. 2009. Mitochondrial nutrients improve immune dysfunction in the type 2 diabetic Goto-Kakizaki rats. *Journal of Cellular and Molecular Medicine* 13:4, 701-711. [CrossRef]
- 32. Nazzareno Ballatori, Suzanne M. Krance, Sylvia Notenboom, Shujie Shi, Kim Tieu, Christine L. Hammond. 2009. Glutathione dysregulation and the etiology and progression of human diseases. *Biological Chemistry* **390**:3, 191-214. [CrossRef]
- 33. Po Sing Leung, Yuk Cheung Chan. 2009. Role of Oxidative Stress in Pancreatic Inflammation. *Antioxidants & Redox Signaling* 11:1, 135-166. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 34. Michael L. Mohler, Yali He, Zhongzhi Wu, Dong Jin Hwang, Duane D. Miller. 2009. Recent and emerging anti-diabetes targets. *Medicinal Research Reviews* **29**:1, 125-195. [CrossRef]
- 35. Consuelo Walss-Bass, Susan T. Weintraub, John Hatch, Jim Mintz, Asish R. Chaudhuri. 2008. Clozapine causes oxidation of proteins involved in energy metabolism: a possible mechanism for antipsychotic-induced metabolic alterations. *The International Journal of Neuropsychopharmacology* 11:08, 1097. [CrossRef]
- 36. John J. Mieyal , Molly M. Gallogly , Suparna Qanungo , Elizabeth A. Sabens , Melissa D. Shelton . 2008. Molecular Mechanisms and Clinical Implications of Reversible Protein S-Glutathionylation. *Antioxidants & Redox Signaling* 10:11, 1941-1988. [Abstract] [Full Text HTML] [Full Text PDF] [Full Text PDF] with Links]
- 37. M.S. Islam, H. Choi. 2008. Chinese Cabbage (Brassica campestris L.) does not Improve Glucose Tolerance, Serum Insulin, or Blood Lipid Profiles in a Rat Model of Type-2 Diabetes. *Journal of Food Science* **73**:9, H213-H217. [CrossRef]
- 38. Brian D. Lamon, David P. Hajjar. 2008. Inflammation at the Molecular Interface of Atherogenesis. *The American Journal of Pathology* **173**:5, 1253-1264. [CrossRef]

- 39. Benbo Song, Donalyn Scheuner, David Ron, Subramaniam Pennathur, Randal J. Kaufman. 2008. Chop deletion reduces oxidative stress, improves # cell function, and promotes cell survival in multiple mouse models of diabetes. *Journal of Clinical Investigation* 118:10, 3378-3389. [CrossRef]
- 40. Orit Rozenberg, Maayan Shiner, Michael Aviram, Tony Hayek. 2008. Paraoxonase 1 (PON1) attenuates diabetes development in mice through its antioxidative properties. *Free Radical Biology and Medicine* **44**:11, 1951-1959. [CrossRef]
- 41. Emiko Manabe, Osamu Handa, Yuji Naito, Katsura Mizushima, Satomi Akagiri, Satoko Adachi, Tomohisa Takagi, Satoshi Kokura, Takashi Maoka, Toshikazu Yoshikawa. 2008. Astaxanthin protects mesangial cells from hyperglycemia-induced oxidative signaling. *Journal of Cellular Biochemistry* **103**:6, 1925-1937. [CrossRef]
- 42. Motoo Yamauchi, Hiroshi Kimura. 2008. Oxidative Stress in Obstructive Sleep Apnea: Putative Pathways to the Cardiovascular Complications. *Antioxidants & Redox Signaling* 10:4, 755-768. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 43. Joseph Tibaldi. 2008. Preserving insulin secretion in Type 2 diabetes mellitus. *Expert Review of Endocrinology & Metabolism* 3:2, 147-159. [CrossRef]
- 44. Keith A. Webster. 2008. Stress hyperglycemia and enhanced sensitivity to myocardial infarction. *Current Hypertension Reports* **10**:1, 78-84. [CrossRef]
- 45. Jyoti D. Malhotra, Randal J. Kaufman. 2007. Endoplasmic Reticulum Stress and Oxidative Stress: A Vicious Cycle or a Double-Edged Sword?. *Antioxidants & Redox Signaling* 9:12, 2277-2294. [Abstract] [Full Text PDF] [Full Text PDF with Links]
- 46. Connie Chen, Mehrdad Arjomandi, John Balmes, Ira Tager, Nina Holland. 2007. Effects of Chronic and Acute Ozone Exposure on Lipid Peroxidation and Antioxidant Capacity in Healthy Young Adults. *Environmental Health Perspectives* **115**:12, 1732-1737. [CrossRef]
- 47. Dr. Eiichi Araki, Jun-Ichi Miyazaki. 2007. Metabolic Disorders in Diabetes Mellitus: Impact of Mitochondrial Function and Oxidative Stress on Diabetes and Its Complications. *Antioxidants & Redox Signaling* **9**:3, 289-291. [Citation] [Full Text PDF] [Full Text PDF with Links]